

# Detergent-free purification of membrane proteins using polymer lipid particle (PoLiPa) technology for use in Cryo-EM

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## Introduction

- Our experienced team of protein scientists offer a complete protein production platform, including bespoke construct design, expression in bacterial, insect or mammalian systems and purification of high quality protein in multi-milligram quantities.
- An exciting recent addition to our protein production platform is the polymer-based extraction of membrane proteins into polymer lipid particles (PoLiPas).
- The use of polymer lipid particles avoids many of the problems associated with detergent-based techniques and offers a generic, cost-effective approach to the purification of membrane proteins.
- Using an ion channels and a G-protein coupled receptors (GPCR) as test studies, we showcase our systems for expression and PoLiPa purification of high quality membrane proteins.
- Furthermore, we demonstrate the utility of Cryo-EM for structural analysis of PoLiPa particles.



## Protein Science Platform at Domainex

### Protein science platform includes:

- Bespoke construct design and cloning
- Protein expression services and expression system optimisation using
  - *E. coli*
  - Insect (baculovirus expression system),
  - Mammalian cells (Transient, BacMam or stable cell line generation)
- Purification by affinity, ion-exchange and size-exclusion chromatography

### Protein Characterisation

High quality proteins are a prerequisite for a number of key processes in drug discovery; and naturally our protein production capability pipelines smoothly into other Domainex platforms such as:

- X-ray crystallography and Cryo-EM leading to structure-based drug design
- Assay development
- Biochemical and biophysical screening for Hit identification
- Fragment screening using our *FragmentBuilder* approach

### PoLiPa membrane proteins

- Wide range of expression systems:
- Expression at up to 10L scale
- Range of tags for detection and purification (including His, FLAG, Strep, SNAP, MBP, GST)
- Range of polymers (SMA, DIBMA, SMA-EA, SMI)

## Purification of membrane proteins using PoLiPa

Styrene maleic acid (SMA) and related copolymers form the basis of our PoLiPa platform for detergent free purification of membrane proteins

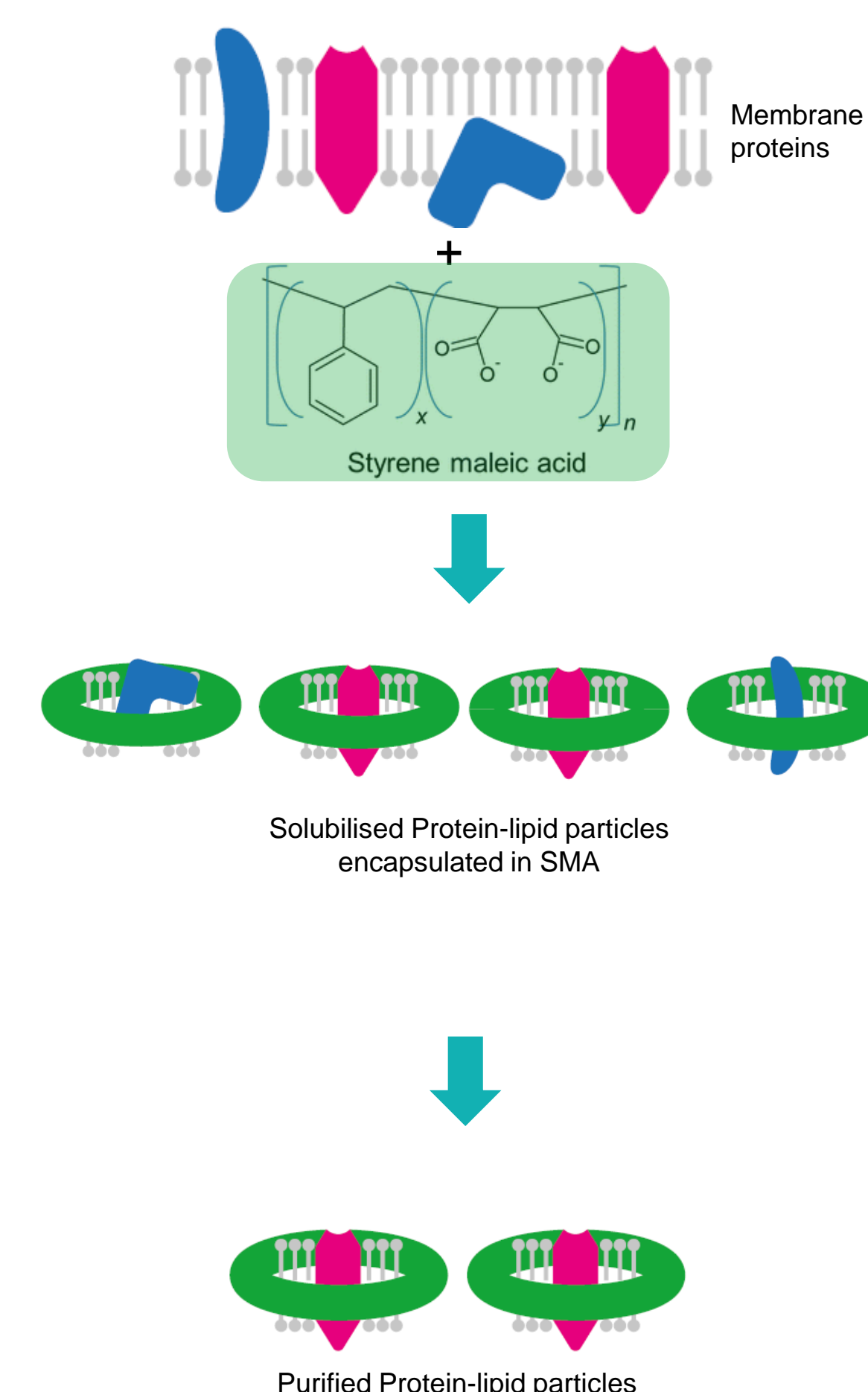
These aliphatic polymers, with a hydrophobic styrene group and a hydrophilic maleic acid group, are able to insert directly into biological membranes and form self-assembling protein/lipid particle

### Solubilisation of lipid membranes

- Polymer (e.g. styrene maleic acid; SMA) inserts into the cell membrane.
- The polymer self-assembles into discs of lipid bilayer with a charged surface that allows solubilisation.
- Membrane proteins contained within these discs will also be solubilised, but remain embedded in their native lipid environment

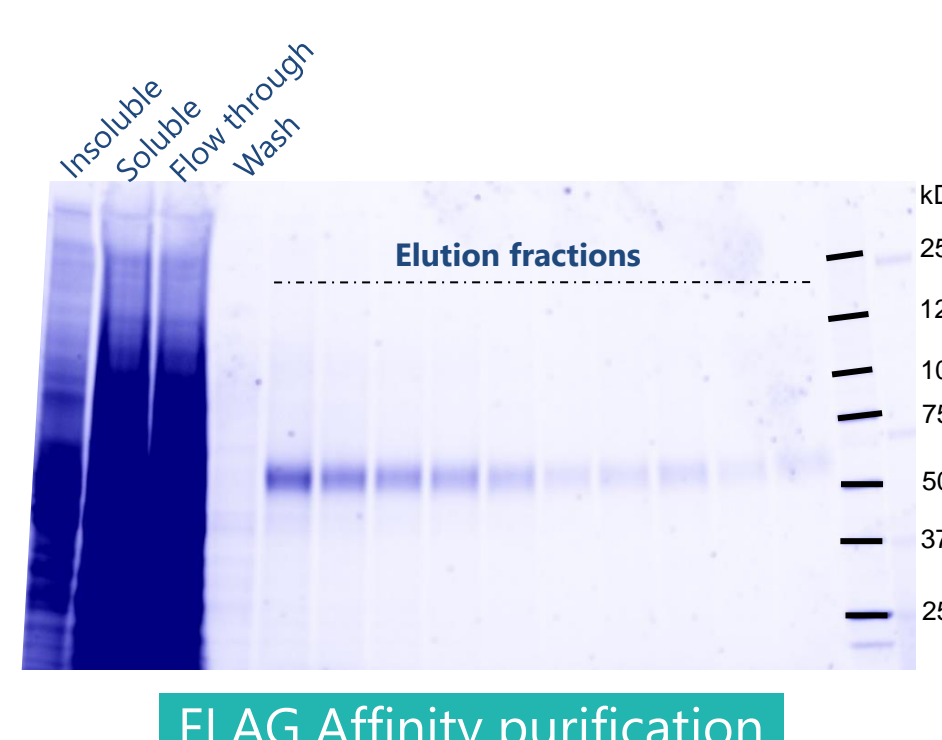
### Purification of proteins using PoLiPa

- Membrane proteins embedded in PoLiPa particles are compatible with conventional chromatography methods (e.g. affinity, size exclusion)
- Generic solubilisation conditions mean similar purification condition can be used for any membrane protein

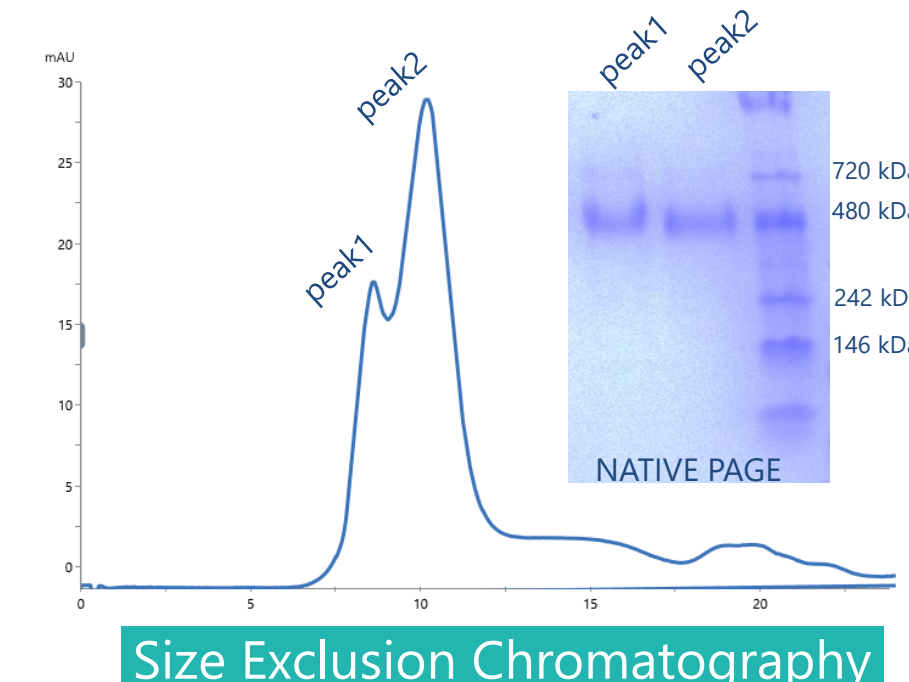


## Case study: Ion channel 1

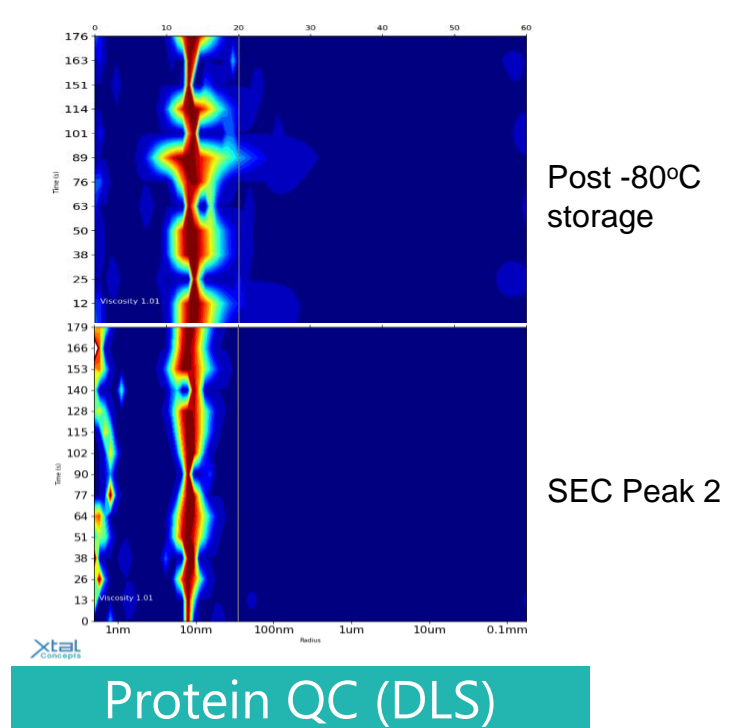
- The membrane fraction from 425ml HEK293 cells, expressing a FLAG-tagged, tetrameric Ca<sup>2+</sup> ion channel protein, was solubilised in 2% SMA before binding to FLAG affinity resin
- In a single step affinity purification, the solubilised ion channel protein was purified to >90% purity



- Subsequent size exclusion chromatography and Native-PAGE analysis demonstrate that the protein can be separated into a single species consistent in size with a tetrameric protein complex + Polipa particle (peak 2)

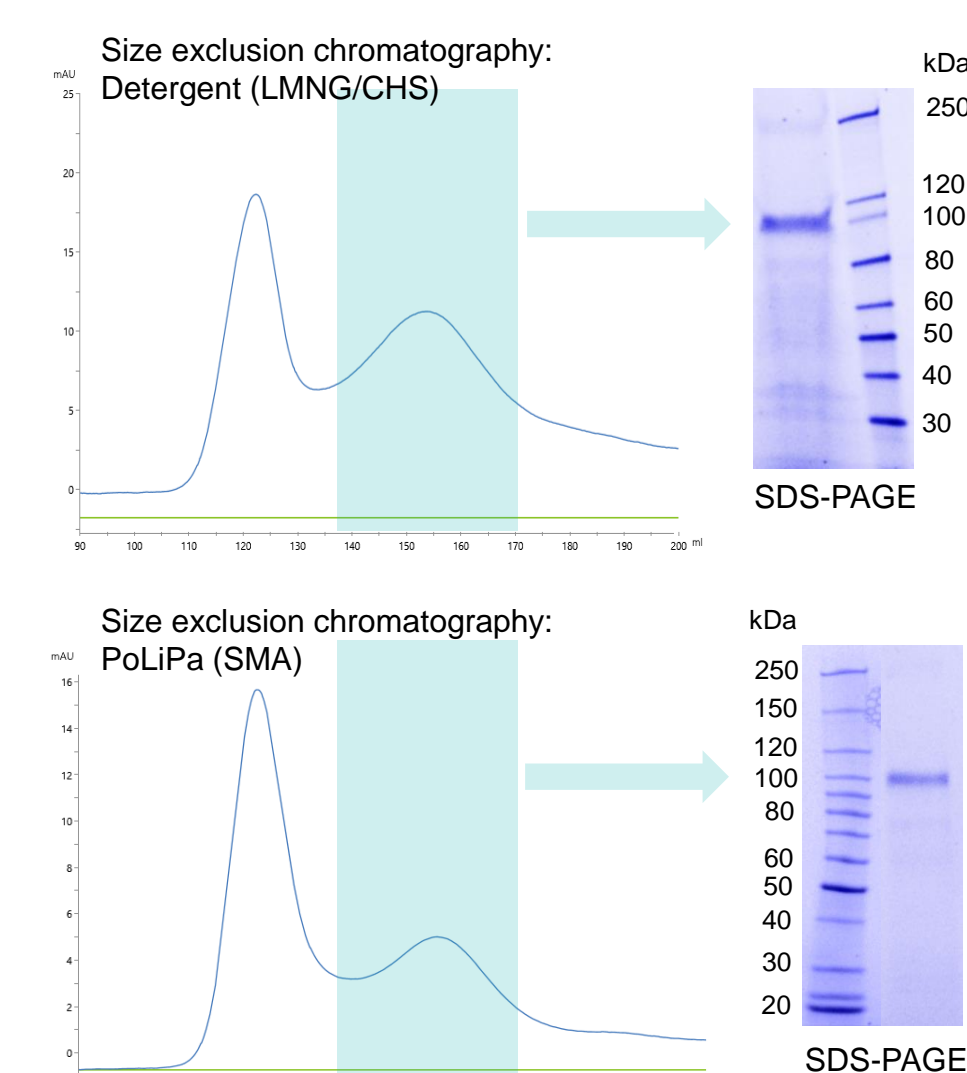


- Dynamic light scattering (DLS) analysis also confirmed the presence of a single population with a diameter of ~10nm, consistent with the known size of SMA lipid particles
- PoLiPa particles are stable over multiple freeze/thaw cycles



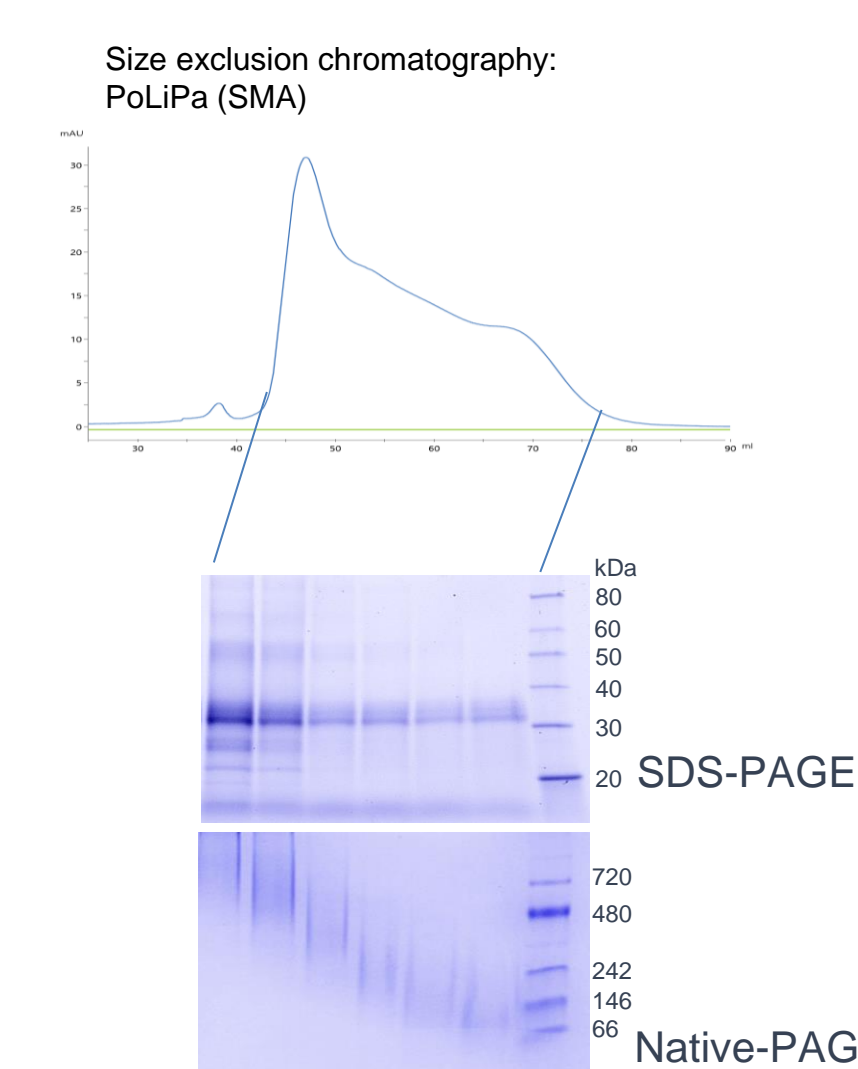
## Case study: Ion channel 2

- A second ion channel protein was expressed in Sf9 insect cells using the baculoviral expression system
- Membrane fractions were obtained and solubilised using either 1% LMNG/CHS or 2% SMA
- Recombinant proteins were purified using Ni-NTA affinity and Size Exclusion chromatography
- Purification yields and size exclusion profiles were comparable for both the detergent and SMA based preparations



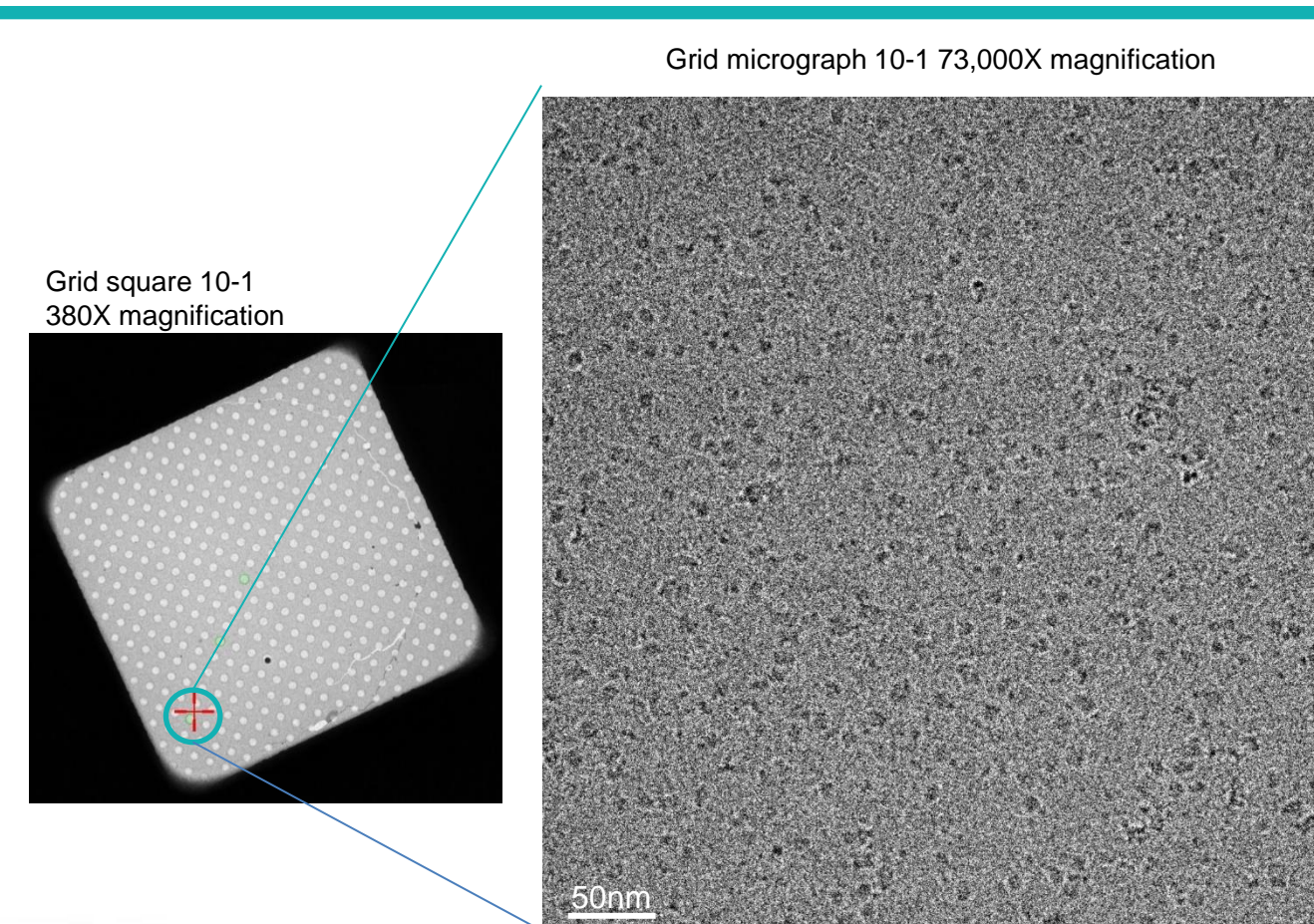
## Case study: GPCR

- A 10xHis tagged GPCR was expressed in Sf21 insect cells using the baculoviral expression system
- Membrane fractions were isolated and solubilised using 2% SMA
- Recombinant GPCR protein was purified using Ni-NTA affinity and Size Exclusion chromatography
- The GPCR could be purified to near homogeneity and, as determined by Native-PAGE, could also be separated into multiple oligomeric species

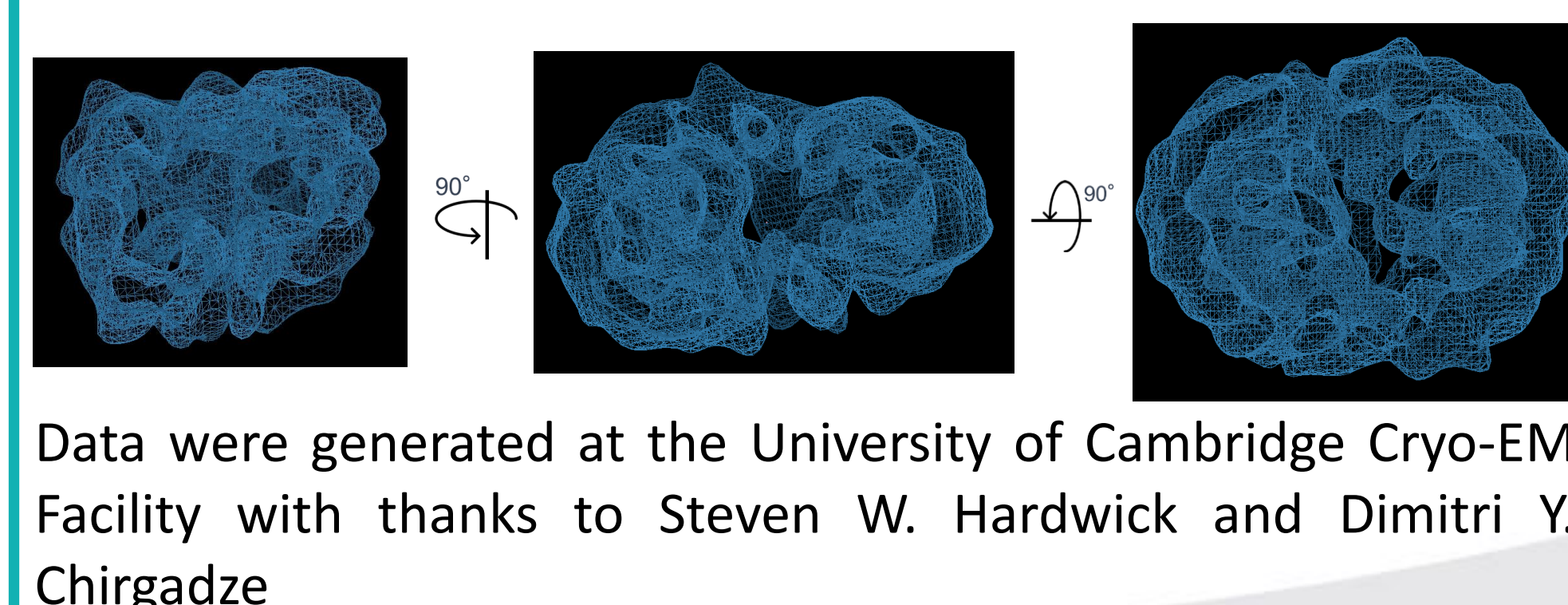


## Cryo-EM grid preparation

- Cryo-EM can be used with many different proteins
- Purified protein is snap frozen into vitrified ice, trapping proteins in multiple orientations
- Protein trapped in the ice is then imaged using an electron microscope
- The images to the right show a grid square with good ice achieved and an example micrograph with a good PoLiPa particle distribution – collected on a Talos Arctica 200Kv



## Cryo-EM PoLiPa 3D map



- A 45kDa protein was used to purify and visualise PoLiPa particles only
- Sufficient particles were picked from a short screening session
- 3D map generation was successful to visualise the PoLiPa disc

## Services/Contact

If you would like to learn more about applying our drug-discovery platforms, please contact: enquiries@domainex.co.uk

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## Conclusions

- Domainex are offering a generic platform approach to solubilise membrane proteins in the complete absence of detergents
- PoLiPa purified protein is amenable to grid generation for Cryo-EM
- These cost-effective and easy to access tools will be invaluable for drug discovery